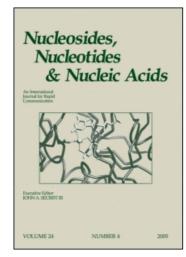
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## Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

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Xiaohua Pengab; Frank Seelaab

<sup>a</sup> Laboratorium für Organische und Bioorganische Chemie, Universität Osnabrück, Osnabrück, Germany <sup>b</sup> Laboratory of Bioorganic Chemistry and Chemical Biology, Center for Nanotechnology, Münster, Germany

To cite this Article Peng, Xiaohua and Seela, Frank(2007) 'An Efficient Synthesis Of 7-Functionalized 7-Deazapurine  $\beta$ -D-Or  $\beta$ -L-Ribonucleosides: Glycosylation Of Pyrrolo[2,3-D]Pyrimidines With 1-O-Acetyl-2,3,5-Tri-O-Benzoyl-D-Or L-Ribofuranose', Nucleosides, Nucleotides and Nucleic Acids, 26: 6, 603 — 606

To link to this Article: DOI: 10.1080/15257770701490332 URL: http://dx.doi.org/10.1080/15257770701490332

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Nucleosides, Nucleotides, and Nucleic Acids, 26:603–606, 2007 Copyright © Taylor & Francis Group, LLC

ISSN: 1525-7770 print / 1532-2335 online DOI: 10.1080/15257770701490332



# AN EFFICIENT SYNTHESIS OF 7-FUNCTIONALIZED 7-DEAZAPURINE $\beta$ -D- OR $\beta$ -L-RIBONUCLEOSIDES: GLYCOSYLATION OF PYRROLO[2,3-d]PYRIMIDINES WITH 1-O-ACETYL-2,3,5-TRI-O-BENZOYL-D- OR L-RIBOFURANOSE

Xiaohua Peng and Frank Seela 

Laboratorium für Organische und Bioorganische Chemie, Universität Osnabrück, Osnabrück, Germany, and Laboratory of Bioorganic Chemistry and Chemical Biology, Center for Nanotechnology, Münster, Germany

☐ The glycosylation reaction performed with 7-halogenated 7-deazapurines employing commercially available 1-O-acetyl-2, 3,5-tri-O-benzoyl-D- or L-ribofuranoses is described.

**Keywords** 7-deazapurines; pyrrolo[2,3-*d*]pyrimidines; glycosylation; functionalization; ribonucleosides; Silyl Hilbert-Johnson reaction

### INTRODUCTION

The frequent occurrence and unusual biological properties of 7-deazapurine nucleosides have promoted studies towards the synthesis, biological activity, and incorporation into oligonucleotides of their chemically designed analogs. The 7-position of 7-deazapurine is an ideal site for modifications that can lead to compounds with increased antiviral activity, oligonucleotides labelled with dyes, or a more stable DNA or RNA duplex through the introduction of substituents of moderate size (e.g., alkynyl residues or halogens). Earlier, 7-deazapurine ribonucleosides (e.g., 1a, 2a, or 3a; Scheme 1) were prepared by nucleobase anion glycosylation using reactive sugar halides. However, until now no efficient procedure has been reported to synthesize 7-functionalized derivatives related to compounds 1a, 2a, or 3a due to the low nucleophilicity of the pyrrol nitrogen in the pyrrolo [2,3-d] pyrimidine system. Here, a convenient method is described for the synthesis of 7-substituted 7-deazapurine

Address correspondence to Frank Seela, Laboratorium für Organische und Bioorganische Chemie, Universität Osnabrück, Barbarastraße 7, 49069 Osnabrück, Germany and Laboratory of Bioorganic Chemistry and Chemical Biology, Center for Nanotechnology, Heisenbergstraße 11, 48149 Münster, Germany. E-mail: frank.seela@uni-osnabrueck.de

**SCHEME 1** Structures of  $\beta$ -D-ribonucleosides 1–3 or  $\beta$ -L-ribonucleosides 4–6.

 $\beta$ -D- or  $\beta$ -L-ribonucleosides (1–3 or 4–6) using commercially available 1-O-acetyl-2,3,5-tri-O-benzoyl-D- or L-ribofuranose (7 or 8).

## RESULTS AND DISCUSSION

The Silyl Hilbert-Johnson glycosylation was employed for the glycosylation of 7-halogenated 2-amino-7-deazapurines with the ribosugar  $7.^{[5,6]}$  The reaction conditions were changed systematically: (i) different nucleobases and silylation reagents (e.g., HMDS and BSA) were employed; (ii) various solvents (CH<sub>2</sub>Cl<sub>2</sub>, ClCH<sub>2</sub>CH<sub>2</sub>Cl, MeNO<sub>2</sub>, and MeCN) were used; (iii) different catalysts (SnCl<sub>4</sub> and TMSOTf) were applied; (iv) different reaction temperatures were tested. When all parameters were carefully chosen, a reliable and convenient procedure was developed which, for the first time, makes 7-functionalized 7-deazapurine ribonucleosides easily accessible. The glycosylation of 2-pivaloylamino-6,7-dihalogenated 7-deazapurines 9b-d with the sugar 7 performed in the one-pot reaction condition (BSA/MeCN/TMSOTf) afforded the  $\beta$ -D-ribonucleosides 10b-d in 73-75% yield (Scheme 2). The intermediates 10b-d are useful for further manipulations using nucleophilic displacement reactions or the palladium-catalyzed cross-coupling reaction leading to a series of

 $\textbf{SCHEME 2} \quad \text{One-pot glycosylation of the nucleobases } \textbf{9b-d} \text{ with sugar derivative } \textbf{7}.$ 

SCHEME 3 Transformation of intermediates 10b-d to the nucleosides 1b-d, 2b-d, and 3b-d.

7-functionalized 7-deazapurine ribonucleosides. This method was also applied to the synthesis of corresponding  $\beta$ -1-ribonucleosides. [5]

The 2,6-diamino ribonucleosides **2b–d** were prepared from **10b–d** when treated with aq. NH<sub>3</sub> (120°C, 24 h) in an autoclave (Scheme 3). The precursors **10b-d** were also converted to the 4-methoxy derivatives **11b–d** (0.5M NaOMe, reflux). The latter were heated in 2M NaOH giving the guanosine analogs **1b–d**. The deamination of the intermediates**11b–d** with NaNO<sub>2</sub>/AcOH furnished the nucleosides **12b–d**, which after demethylation with 2N NaOH gave the 7-deazaxanthosine derivatives **3b-d** (74–78% yield).

The iodinated 7-deazapurine ribonucleosides are valuable starting materials for the introduction of alkynyl or aminoalkynyl chains by the Pdcatalyzed cross-coupling reaction.<sup>[8]</sup> Thus, the 7-iodo compounds **1d**, **2d**, and **3d** were employed in the Sonogashira reaction, yielding a number of

<sup>a</sup> Conditions: (a) HC≡CR, anhydrous DMF, Pd(0)(PPh<sub>3</sub>)<sub>4</sub>, CuI, Et<sub>3</sub>N; (b) MeOH, K<sub>2</sub>CO<sub>3</sub>.

SCHEME 4 Palladium-catalyzed Sonogashira cross-coupling reaction.

**SCHEME 5** Glycosylation of **9b–d** with L-ribofuranose **8**.

novel 7-alkynyl- or aminoalkynyl 7-deazapurine ribonucleosides in 50–92% yields (1e–i, 2e–i or 3e; Scheme 4).

<sup>a</sup> Conditions: (a) HC≡CR, anhydrous DMF, Pd(0) (PPh<sub>3</sub>)<sub>4</sub>, CuI, Et<sub>3</sub>N; (b) MeOH, K<sub>2</sub>CO<sub>3</sub>.

Considering the identical chemical properties of L- and D-nucleosides and their precursors in a nonchiral environment, the protocols developed for the 7-deazapurine D-ribonucleoside synthesis (as described above) can be employed for the preparation of the L-enantiomers. Instead of D-ribofuranose derivative **7**, its L-enantiomer **8** was used as sugar component. The glycosylation of the 2-pivaloylamino-7-deazapurine derivatives **9b–d** with **8** was performed in MeCN/BSA/TMSOTf, affording the  $\beta$ -L-ribonucleosides **13b–d** in 66–73% yield (Scheme 5). [9] Similar to the transformation of **10b–d** into **1–3**, compounds **13b–d** were converted to the 7-functionalized 7-deazapurine  $\beta$ -L-ribonucleosides **4b–d,5b–d**, and**6b–d** by nucleophilic displacement reactions (Scheme 1).

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